

# Mitochondrial Complex I, H<sub>2</sub>O<sub>2</sub> and NO as Prodromal Signals of Cardiac Dysfunction in Type 1 Diabetes

*Complejo I, H<sub>2</sub>O<sub>2</sub> y NO mitocondriales como señales prodrómicas de la disfunción cardíaca en diabetes tipo 1*

IVANA A. RUKAVINA-MIKUSIC<sup>1,2</sup>, MICAELA REY<sup>1</sup>, VALERIA TRÍPODI<sup>3</sup>, LAURA B. VALDEZ<sup>1,2</sup>.

## ABSTRACT

**Background:** Previous results from our laboratory suggest that heart mitochondrial dysfunction precedes myocardial failure associated with sustained hyperglycemia.

**Purpose:** The aim of this study was to analyze the early events that take place in heart mitochondria in a type 1 diabetes mellitus (DM) model.

**Methods:** Male Wistar rats were injected with streptozotocin (STZ; 60 mg/kg, ip.) to induce DM. They were euthanized 10 or 14 days later and the heart mitochondrial fraction was obtained.

**Results:** State 3 O<sub>2</sub> consumption in the presence of malate-glutamate (21%) or succinate (16%), and complex I-III (27%), II-III (24%) and IV (22%) activities were lower in diabetic animals 14 days after STZ injection. When animals were euthanized at day 10, only state 3 O<sub>2</sub> consumption sustained by complex I substrates (23%) and its corresponding respiratory control (30%) were lower in rats injected with STZ, in agreement with reduced complex I-III activity (17%). These changes were accompanied by increased H<sub>2</sub>O<sub>2</sub> (117%), NO (30%) and ONOO<sup>-</sup> (~225%) production rates, mtNOS expression (29%) and O<sub>2</sub><sup>-</sup> (~150%) and NO (~30%) steady-state concentrations, together with a decrease in Mn-SOD activity (15%) and mitochondrial [GSSG+GSH] (28%), without changes in PGC-1α expression.

**Conclusion:** Complex I dysfunction and increased H<sub>2</sub>O<sub>2</sub>, NO and ONOO<sup>-</sup> production rates can be considered subcellular prodromal signals of the mitochondrial damage that precedes myocardial dysfunction in diabetes.

**Key Words:** Type 1 Diabetes Mellitus - Hyperglycemia - Mitochondrial Complex I - Heart mitochondria - Nitric Oxide - Hydrogen Peroxide

## RESUMEN

**Introducción:** Resultados del laboratorio sugieren que la disfunción mitocondrial en el corazón precede a la falla miocárdica asociada a la hiperglucemia sostenida.

**Objetivo:** Estudiar los eventos tempranos que ocurren en las mitocondrias de corazón en un modelo de diabetes mellitus (DM) tipo 1.

**Materiales y métodos:** Ratas Wistar macho fueron inyectadas con estreptozotocina (STZ; 60 mg/kg, ip) y sacrificadas 10 o 14 días posinyección. Se obtuvo la fracción mitocondrial de corazón.

**Resultados:** El consumo de O<sub>2</sub> en estado 3 en presencia de malato-glutamato (21%) o succinato (16%) y las actividades de los complejos I-III (27%), II-III (24%) y IV (22%) fueron menores en los animales diabéticos a los 14 días posinyección. Cuando los animales se sacrificaron al día 10, solo el consumo de O<sub>2</sub> en estado 3 en presencia de sustratos del complejo I (23%) y su control respiratorio (30%) fueron menores en las ratas inyectadas con STZ, de acuerdo con una reducción en la actividad del complejo I-III (17%). Estos cambios se acompañaron de un aumento en las velocidades de producción de H<sub>2</sub>O<sub>2</sub> (117%), NO (30%) y ONOO<sup>-</sup> (~225%), en la expresión de mtNOS (29%) y en la [O<sub>2</sub><sup>-</sup>]<sub>ss</sub> (~150%) y [NO]<sub>ss</sub> (~30%), junto con una disminución de la actividad de la Mn-SOD (15%) y la [GSSG+GSH] mitocondrial (28%), sin cambios en la expresión de PGC-1α.

**Conclusión:** La disfunción del complejo I y el aumento en la generación de H<sub>2</sub>O<sub>2</sub>, NO y ONOO<sup>-</sup> pueden considerarse señales subcelulares prodrómicas del deterioro de la función mitocondrial que precede a la disfunción cardíaca en la diabetes.

**Palabras clave:** Diabetes mellitus tipo 1 - Hiperglucemia - Complejo I mitocondrial - Mitocondrias cardíacas - Óxido nítrico - Peróxido de hidrógeno

REV ARGENT CARDIOL 2021;89:90-95. <http://dx.doi.org/10.7775/rac.v89.i2.19865>

Received: 12/14/2020 – Accepted: 02/14/2021

**Address for reprints:** Dra. Laura B. Valdez - Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Departamento de Química Analítica y Físicoquímica - Junín 956, C1113AAD, Buenos Aires, Argentina - Tel: 54-11-5287-4235 - E-mail: lbvaldez@ffyb.uba.ar; laubeaval@gmail.com

<sup>1</sup> Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Departamento de Química Analítica y Físicoquímica, Físicoquímica;

<sup>2</sup> Consejo Nacional de Investigaciones Científicas y Técnicas, Instituto de Bioquímica y Medicina Molecular (IBIMOL; UBA-CONICET), Físicoquímica

<sup>3</sup> Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Departamento de Tecnología Farmacéutica. Buenos Aires, Argentina.

## INTRODUCTION

Diabetes Mellitus is a chronic metabolic disease characterized by an increase in blood glucose levels, which could be developed as a result of defects in insulin secretion, insulin action, or both. Cardiovascular complications are the main cause of morbidity and mortality among the diabetic population (1, 2), with sustained hyperglycemia being one of the risk factors that ultimately leads to diabetic cardiomyopathy (3, 4). Although the latter is frequently associated with type 2 diabetes, patients with type 1 diabetes also suffer a metabolically induced cardiomyopathy, which occurs independently of hypertension, nephropathy, or ischemic heart disease (5, 6). Previous results from our laboratory, obtained using an experimental model of type 1 diabetes in rat, show that the sustained hyperglycemia for 25 days leads to generalized cardiac mitochondrial dysfunction, in the absence of hypertrophy and changes in resting cardiac function, but with cardiac compromise against a work overload (7, 8), suggesting that mitochondrial function impairment precedes myocardial failure in diabetic patients.

Considering that most of the published studies refer to cardiac mitochondrial function in models of type 2 diabetes—in association with insulin resistance, obesity, hypercholesterolemia and hypertension—or in type 1 diabetic models during very prolonged hyperglycemia, the purpose of this work was to study the early events that occur in rat heart mitochondria in an experimental model of type 1 diabetes.

## METHODS

Male Wistar rats (200–250 g) were randomly divided into two groups: control (C) and diabetes mellitus (DM). Diabetes was induced by an intraperitoneal injection of streptozotocin (STZ, 60 mg/kg) diluted in citrate buffer 100 mM pH 4.50. Group C animals received an intraperitoneal injection of the vehicle (citrate buffer 100 mM pH 4.50). Glycemia was determined 3 days after STZ injection and throughout the experimental protocol, using a One Touch Ultra, Johnson & Johnson glucometer. Rats with blood glucose concentration >200 mg/dl 3 days after STZ injection were considered diabetic (9). Animal weight was assessed throughout the study period. At 10 or 14 days post-STZ injection, the animals were euthanized in a CO<sub>2</sub> atmosphere, the heart was removed, and the mitochondrial fraction was obtained by differential centrifugation (10).

Mitochondrial O<sub>2</sub> consumption was assessed in the presence of complex I (malate plus glutamate) or complex II (succinate) substrates, both in the absence (state 4 or passive respiration) as in the presence (state 3 or active respiration) of ADP using a Clark electrode (Hansatech Oxygraph, Hansatech Instruments Ltd, Norfolk, England) (11). Respiratory control (RC) was calculated as state 3/state 4 O<sub>2</sub> consumption ratio. The respiratory complex I-III (NADH-cytochrome c reductase), II-III (succinate-cytochrome c reductase) and IV (cytochrome oxidase) activities were assessed at 550–540 nm with a diode array spectrophotometer (12).

The mitochondrial hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) production rate was measured fluorometrically at 365–450 nm using the scopoletin-horseradish peroxidase (HRP) method,

in the presence of complex I substrates (malate plus glutamate) (12, 13). Mitochondrial nitric oxide (NO) production was determined through the oxidation of oxyhemoglobin to methemoglobin with a diode array spectrophotometer, at 577–591 nm (12, 14, 15). Mn-superoxide dismutase (Mn-SOD) activity was assessed spectrophotometrically from the ferricytochrome c to ferrocyanochrome c reduction rate by the superoxide anion (O<sub>2</sub><sup>•-</sup>) using the xanthine-xanthine oxidase system (16). Steady state O<sub>2</sub><sup>•-</sup> and NO concentrations ([O<sub>2</sub><sup>•-</sup>]ss and [NO]ss) were calculated in the mitochondrial matrix (8) from the experimental measurements of H<sub>2</sub>O<sub>2</sub>, NO and Mn-SOD, and the peroxynitrite (ONOO<sup>-</sup>) production rate was estimated from these concentrations. Mitochondrial concentrations of reduced (GSH) and oxidized (GSSG) glutathione were determined by micro HPLC-MS/MS (17). Mitochondrial nitric oxide synthase (mtNOS) and peroxisomal proliferation-activated receptor  $\gamma$  coactivator 1 $\alpha$  (PGC-1 $\alpha$ ) expressions were analyzed by western blot using neuronal anti-NOS (H299: sc-8309, Santa Cruz Biotechnology) and anti-PGC-1 $\alpha$  (K-15: sc-5816, Santa Cruz Biotechnology) antibodies, respectively. Anti-VDAC-1 (D-16: sc-32063, Biotechnology) and anti- $\beta$ -tubulin ([1E1-E8-H4] ab131205, Abcam) antibodies were used as loading controls.

## Statistical analysis

The results included in tables and figures are expressed as mean  $\pm$  SEM and represent repetitions of 3 to 10 independent experiments. Statistical analysis was performed using the GraphPad InStat 4 program (GraphPad Software, La Jolla, CA, USA). Student's t test was used to analyze significant differences between C and DM groups, and ANOVA-Dunnett's test to analyze the significant differences of group C or group DM with respect to treatment onset (day 0).

## Ethical considerations

The experimental protocol was approved by the Institutional Committee for the Care and Use of Laboratory Animals of the School of Pharmacy and Biochemistry of University of Buenos Aires (CICUAL; 00663658/15) and the procedures were carried out following the indications of the Guide for the Care and Use of Laboratory Animals published by the United States National Academy of Sciences.

## RESULTS

Glycemia assessed on the third day after STZ injection was significantly higher in diabetic animals compared with the C group and remained elevated throughout the experimental period, with no differences within the DM group throughout the treatment period (Fig. 1A).

The weight of the heart of STZ-treated animals at 10 days and 14 days post-injection was 15% and 25% lower respectively compared with the weight of control animals, in agreement with the lower increase in body weight observed in diabetic rats with respect to the normal growth profile of the C group (Fig. 1B). Consequently, there were no differences in the cardiac index (heart weight/body weight ratio) between the groups (C: 0.340  $\pm$  0.010; DM-10 days: 0.336  $\pm$  0.004; DM-14 days: 0.339  $\pm$  0.003), indicating the absence of hypertrophy at these experimental times.

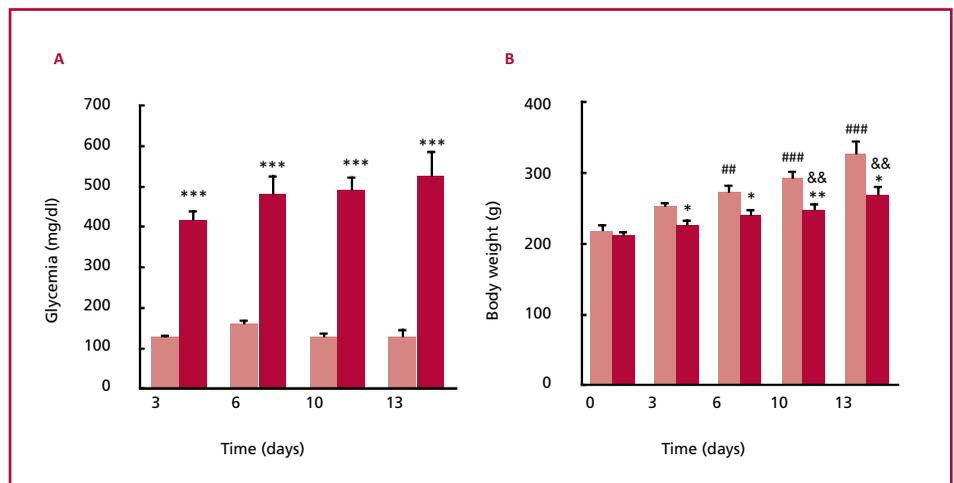
In order to evaluate the cardiac mitochondrial function, O<sub>2</sub> consumption in the presence of malate-glutamate or succinate, as well as respiratory complex-

es I-III, II-III and IV activities were measured (Table 1). No changes in state 4 (passive) mitochondrial respiration, sustained by malate-glutamate or succinate, were observed in the heart of diabetic rats compared with control animals, neither at 10 nor at 14 days after STZ injection. When the rats were euthanized on day 10, which corresponds to approximately 7 days of hyperglycemia, state 3 O<sub>2</sub> consumption (active respiration), sustained by malate-glutamate (23%), as well as its corresponding respiratory control (30%), were lower in the diabetic group, in agreement with the reduction in complex I-III (17%) activity, with complex I being the only modified respiratory complex at this stage. On the contrary, when the animals were euthanized 14 days after STZ injection, that is, after ~11 days of hyperglycemia, both state 3 O<sub>2</sub> consumption, sustained by malate-glutamate (21%) as state 3 respiration, sustained by succinate (16%), and respiratory complex I-III (27%), II-III (24%) and IV (22%) activities were lower in the diabetic group in relation to the values detected in the control group.

Since ~7 days of hyperglycemia led to an incipient mitochondrial dysfunction, evidenced both by the decrease in O<sub>2</sub> consumption in state 3 sustained by malate-glutamate as by the reduction of complex I-III activity, that experimental period was considered an early stage of cardiac mitochondrial dysfunction. For this reason, reactive oxygen and nitrogen species generation, the redox state and mitochondrial biogenesis were studied in the heart of animals euthanized 10 days after STZ injection.

The mitochondrial redox state was evaluated by measuring the concentration of mitochondrial glutathione and Mn-SOD activity, the main mitochondrial non-enzymatic and enzymatic antioxidants, respectively (Table 2). The mitochondrial concentration of total glutathione ([GSH+GSSG]) was 28% lower in diabetic animals compared with that determined in control rats, due both to a decrease in the concentration of reduced glutathione ([GSH]) as in the concentration of oxidized glutathione ([GSSG]), with no differences in the [GSH]/[GSSG] ratio. Mn-SOD activity was also

**Fig. 1.** Glycemia (A) and body weight (B) of the animals throughout the experimental protocol. Group C: pink bars; group DM: red bars. \*\*\* p <0.005; \*\* p <0.01; \* p <0.05 DM vs. C on the same treatment day (Student's t test); ### p <0.005; ## p <0.01 C vs. C at day 0 (ANOVA-Dunnett); && p <0.01 DM vs. DM at day 0 (ANOVA-Dunnett).



**Table 1.** Mitochondrial function. O<sub>2</sub> consumption and respiratory complexes in heart mitochondria of control and diabetic rats, euthanized 10 or 14 days after STZ injection. C: control, DM: diabetes mellitus

	C	DM	DM
		10 days	14 days
<b>MITOCHONDRIAL O<sub>2</sub> CONSUMPTION</b>			
<b>MALATE-GLUTAMATE</b>			
State 4 (ng-at. O/min.mg protein)	41 ± 3	45 ± 4	39 ± 3
State 3 (ng-at. O/min.mg protein)	353 ± 20	271 ± 24*	280 ± 18*
Respiratory control	8.6	6.0	7.2
<b>SUCCINATE</b>			
State 4 (ng-at. O/min.mg protein)	122 ± 5	126 ± 11	106 ± 13
State 3 (ng-at. O/min.mg protein)	347 ± 21	299 ± 23	290 ± 30
Respiratory control	2.8	2.4	2.7
<b>RESPIRATORY COMPLEXES</b>			
I-III (nmol/min.mg protein)	376 ± 14	312 ± 18*	274 ± 26***
II-III (nmol/min.mg protein)	184 ± 10	154 ± 16	140 ± 13*
IV (1/min.mg protein)	72 ± 4	69 ± 5	56 ± 4*

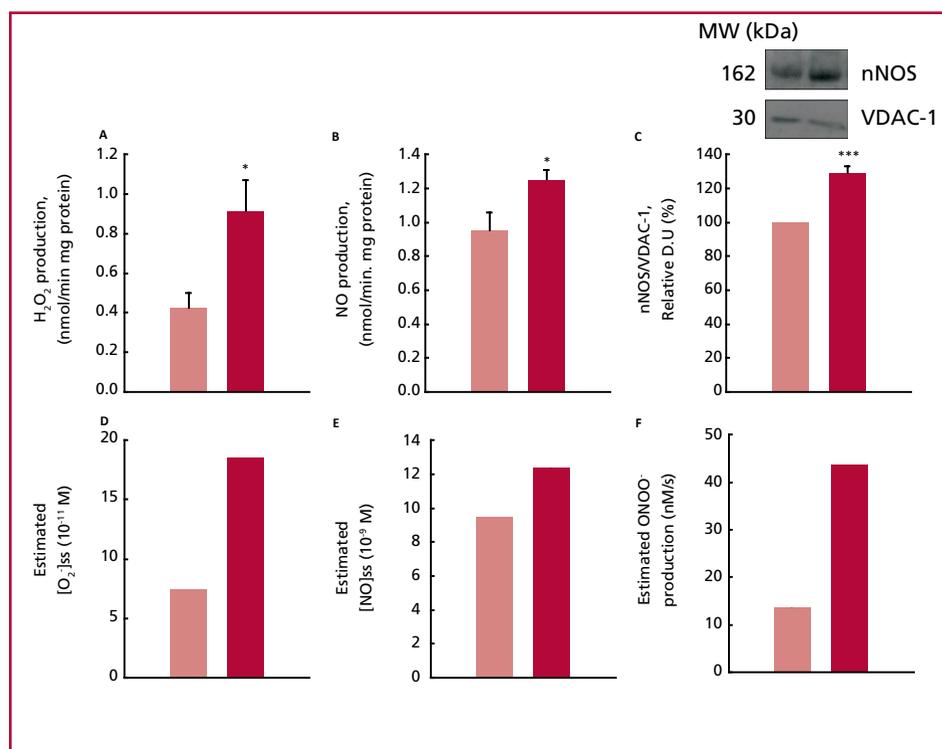
\*\*\*p<0.005; \*p<0.05 DM vs. C (Student's t test)

	C	DM - 10 days
<b>[Glutathione] (pmol/mg protein)</b>		
[GSH]	104 ± 5	76 ± 7
[GSSG]	8 ± 1	5.5 ± 0.7
[GSH+GSSG]	113 ± 6	81 ± 7*
[GSH]/[GSSG]	12 ± 1	14 ± 2
<b>Mn-SOD</b>		
Activity (USOD/mg protein)	79 ± 5	67.5 ± 2.5*
[Mn-SOD] (µM)	11.2 ± 0.8	9.8 ± 0.4

\*  $p < 0.05$  DM vs. C (Student's t test)

**Table 2.** Mitochondrial redox state. [GSH], [GSSG] and Mn-SOD in hearts mitochondria of control and diabetic rats euthanized 10 days after STZ injection.

GSH: reduced glutathione  
GSSG: oxidized glutathione  
Mn-SOD: Manganese superoxide dismutase



**Fig. 2.** Reactive oxygen and nitrogen species. Mitochondrial H<sub>2</sub>O<sub>2</sub>-malate+glutamate- (A), NO (B) and ONOO<sup>-</sup> production (F). mtNOS expression (C). Steady-state mitochondrial O<sub>2</sub><sup>-</sup> (D) and NO concentrations (E). C group: pink bars; DM group: red bars.

\*\*\*  $p < 0.005$ ; \*  $p < 0.05$  DM vs. C (Student's t test)

H<sub>2</sub>O<sub>2</sub>: hydrogen peroxide  
NO: nitric oxide ONOO<sup>-</sup>: peroxynitrite

found to be 15% lower in the heart mitochondria of diabetic animals euthanized 10 days after STZ injection.

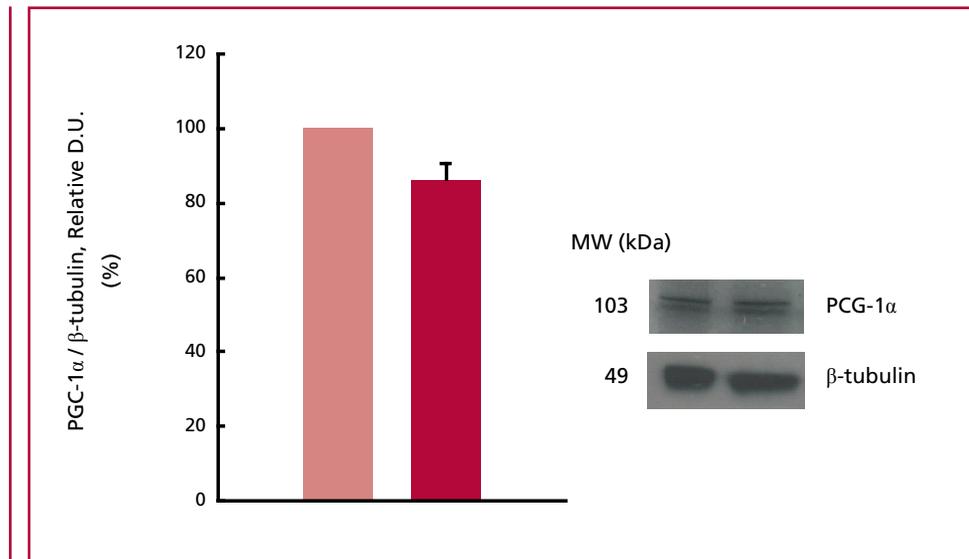
Regarding the production rates of reactive oxygen species, the generation of H<sub>2</sub>O<sub>2</sub> in the presence of complex I substrates was significantly higher (117%) in the heart mitochondria of diabetic animals after ~7 days of hyperglycemia, compared with that assessed in the control group (Fig. 2A). The rate of mitochondrial NO generation also increased by 30% (Fig. 2B), in agreement with 29% increase in the expression of NOS in mitochondrial membranes (mtNOS) (Fig. 2C).

The steady-state concentrations of O<sub>2</sub><sup>-</sup>—stoichiometric precursor of H<sub>2</sub>O<sub>2</sub>— and NO were estimated from experimental measurements that have been performed in the heart mitochondria of diabetic animals and controls. A very marked increase (~150%) in [O<sub>2</sub>]<sup>ss</sup> (Fig. 2D) was observed in the diabetic group compared with the control group, together with ~30% increase in [NO]<sup>ss</sup> (Fig. 2E). The rate of mitochondrial

ONOO<sup>-</sup> generation (Fig. 2F) estimated from the steady state concentrations of these species, resulted 3 times higher in the heart of animals injected with STZ than in control rats.

Moreover, PGC-1 $\alpha$  transcription factor expression, the main regulator of mitochondrial biogenesis, was determined in total heart homogenate from animals euthanized 10 days after STZ injection, and cardiac mitochondrial mass was estimated from the ratio of cytochrome oxidase (COX) activity assessed in the mitochondrial fraction and in the homogenate (18). No differences were found between the experimental groups, neither in PGC-1 $\alpha$  expression (Fig. 3) nor in the cardiac mitochondrial mass (C: 24.6 ± 0.1 and DM: 24.1 ± 0.2 mg mitochondrial protein/g tissue), indicating that after 7 days of hyperglycemia, de novo mitochondrial synthesis has not been triggered, contrary to what happens when hyperglycemia is sustained for 25 days (8).

**Fig. 3.** Mitochondrial biogenesis. PGC-1 $\alpha$  expression in heart homogenate from control and diabetic rats. Group C: pink bars; group DM: red bars. PGC-1  $\alpha$ : peroxisomal proliferation-activated receptor gamma coactivator 1  $\alpha$



## DISCUSSION

The results show that ~7 days of sustained hyperglycemia are sufficient to show incipient cardiac mitochondrial dysfunction. The decrease in state 3  $O_2$  consumption and the enhancement in  $H_2O_2$  production, in both cases when using malate-glutamate, in addition with the reduction of complex I activity, suggest the presence of structural modifications of complex I proteins in the first phase of cardiac dysfunction in diabetic rats. The increase in  $H_2O_2$  production was accompanied by an increase in ONOO $\cdot$  generation, the latter at the expense of a marked increment in  $O_2\cdot^-$  concentration and in NO generation and mtNOS expression, the enzyme responsible for its synthesis. It should be mentioned that ONOO $\cdot$  is a highly oxidizing and nitrating species, involved in complex I inhibition through tyrosine nitration and/or nitrosylation. (12, 19). Since no changes were evidenced in PGC-1 $\alpha$  factor expression or in cardiac mitochondrial mass, mitochondrial NO seems to be a chemical species located upstream in the signaling pathways that lead to the synthesis of new mitochondria in response to hyperglycemia.

In conclusion, complex I dysfunction and increased mitochondrial production rates of  $H_2O_2$ , NO, and ONOO $\cdot$  can be considered subcellular prodromal signals of the cardiac mitochondrial dysfunction that precedes myocardial failure in this type 1 DM model.

## Conflicts of interest

None declared.

(See authors' conflicts of interest forms on the website/ Supplementary material)

## Funding:

This work was carried out with research grants from Universidad de Buenos Aires (UBACyT 200-201-601-00132BA)

and from Agencia Nacional de Promoción Científica y Tecnológica (ANPCyT, PICT 2016-0187).

## REFERENCES

- Garcia MJ, McNamara PM, Gordon T, Kannell WB. Morbidity and mortality in diabetics in the Framingham population. Sixteen year follow up study. *Diabetes* 1974;23:105–11. <https://doi.org/10.2337/diab.23.2.105>
- Miki T, Yuda S, Kouzu H, Miura T. Diabetic cardiomyopathy: Pathophysiology and clinical features. *Heart Fail. Rev* 2013; 18:149–66. <https://doi.org/10.1007/s10741-012-9313-3>
- Boudina S, Abel ED. Diabetic cardiomyopathy revisited. *Circulation* 2007;115:3213–23. <https://doi.org/10.1161/CIRCULATIONAHA.106.679597>
- Huynh K, Bernardo BC, McMullen JR, Ritchie RH. Diabetic cardiomyopathy: Mechanisms and new treatment strategies targeting antioxidant signaling pathways. *Pharmacol Ther* 2014;142:375–415. <https://doi.org/10.1016/j.pharmthera.2014.01.003>
- Götzsche O, Darwish A, Göttsche L, Hansen LP, Sørensen KE. Incipient Cardiomyopathy in Young Insulin-dependent Diabetic Patients: A Seven-year Prospective Doppler Echocardiographic Study. *Diabet Med* 1996;13:834–40. [https://doi.org/10.1002/\(SICI\)1096-9136\(199609\)13:9<834::AID-DIA225>3.0.CO;2-M](https://doi.org/10.1002/(SICI)1096-9136(199609)13:9<834::AID-DIA225>3.0.CO;2-M)
- Ritchie RH, Zerenturk EJ, Prakoso D, Calkin AC. Lipid metabolism and its implications for type 1 diabetes-associated cardiomyopathy. *J Mol Endocrinol* 2017;58:R225–40. <https://doi.org/10.1530/JME-16-0249>
- Bombicino SS, Iglesias DE, Mikusic IAR, D'Annunzio V, Gelpi RJ, Boveris A, et al. Diabetes impairs heart mitochondrial function without changes in resting cardiac performance. *Int. J Biochem Cell Biol* 2016;81:335–45. <https://doi.org/10.1016/j.biocel.2016.09.018>
- Bombicino SS, Iglesias DE, Rukavina-Mikusic IA, Buchholz B, Gelpi RJ, Boveris A, et al. Hydrogen peroxide, nitric oxide and ATP are molecules involved in cardiac mitochondrial biogenesis in Diabetes. *Free Radic Biol. Med* 2017;112:267–76. <https://doi.org/10.1016/j.freeradbiomed.2017.07.027>
- Joffe II, Travers KE, Perreault-Micale CL, Hampton T, Katz SE, Morgan JP, et al. Abnormal cardiac function in the streptozotocin-induced non-insulin-dependent diabetic rat: noninvasive assessment with doppler echocardiography and contribution of the nitric oxide pathway. *J Am Coll Cardiol* 1999;34:2111–9. [https://doi.org/10.1016/S0735-1097\(99\)00436-2](https://doi.org/10.1016/S0735-1097(99)00436-2)
- Boveris A, Oshino N, Chance B. The cellular production of hydrogen peroxide. *Biochem J* 1972;128:617–30. <https://doi.org/10.1042/bj1280617>
- Boveris A, Costa LE, Cadenas E, Poderoso JJ. Regulation of

- mitochondrial respiration by adenosine diphosphate, oxygen and nitric oxide, *Methods Enzymol* 1999;301:188–98. [https://doi.org/10.1016/S0076-6879\(99\)01082-4](https://doi.org/10.1016/S0076-6879(99)01082-4)
12. Valdez LB, Zaobornyj T, Bombicino S, Iglesias DE, Boveris A, Donato M, et al. Complex I syndrome in myocardial stunning and the effect of adenosine. *Free Radic Biol Med* 2011;51:1203–12. <https://doi.org/10.1016/j.freeradbiomed.2011.06.007>
13. Boveris A. Determination of the production of superoxide radicals and hydrogen peroxide in mitochondria, *Methods Enzymol* 1984;105:429–35. [https://doi.org/10.1016/S0076-6879\(84\)05060-6](https://doi.org/10.1016/S0076-6879(84)05060-6)
14. Murphy M, Noack E. Nitric oxide assay using the hemoglobin method, *Methods Enzymol* 1994; 233:240–50. [https://doi.org/10.1016/S0076-6879\(94\)33027-1](https://doi.org/10.1016/S0076-6879(94)33027-1)
15. Boveris A, Lores Arnaiz S, Bustamante J, Alvarez S, Valdez LB, Boveris AD, et al. Pharmacological regulation of mitochondrial nitric oxide synthase, *Methods Enzymol* 2002; 359:328–39. [https://doi.org/10.1016/S0076-6879\(02\)59196-5](https://doi.org/10.1016/S0076-6879(02)59196-5)
16. McCord JM, Fridovich I. Superoxide dismutase. An enzymic function for erythrocyte hemoglobin, *J Biol Chem* 1969;244:6049–55. [https://doi.org/10.1016/S0021-9258\(18\)63504-5](https://doi.org/10.1016/S0021-9258(18)63504-5)
17. Squellerio I, Caruso D, Porro B, Veglia F, Tremoli E, Cavalca V. Direct glutathione quantification in human blood by LC-MS/MS: comparison with HPLC with electrochemical detection, *J Pharm Biomed Anal* 2012;71:111–8. <https://doi.org/10.1016/j.jpba.2012.08.013>
18. Navarro A, Bandez MJ, Lopez-Cepero JM, Gómez C, Boveris AD, Cadenas E, et al. High doses of vitamin E improve mitochondrial dysfunction in rat hippocampus and frontal cortex upon aging, *Am. J Physiol Regul Integr Comp Physiol* 2011;300:R827–R834. <https://doi.org/10.1152/ajpregu.00525.2010>
19. Riobó NA, Clementi E, Melani M, Boveris A, Cadenas E, Moncada S, et al. Nitric oxide inhibits mitochondrial NADH:ubiquinone reductase activity through peroxynitrite formation. *Biochem J* 2001;359:139–45. <https://doi.org/10.1042/bj3590139>